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Dalton Utilities LAS Carpet company overapplying spraying to a creek which ended up in the river. 10 yrs ago. Unsure if a consent decree came out of it.



# WEISS LAKE PRIORITIZATION

**May 2019**

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# **PRIORITIZATION OF PFAS CONTRIBUTIONS TO WEISS LAKE**

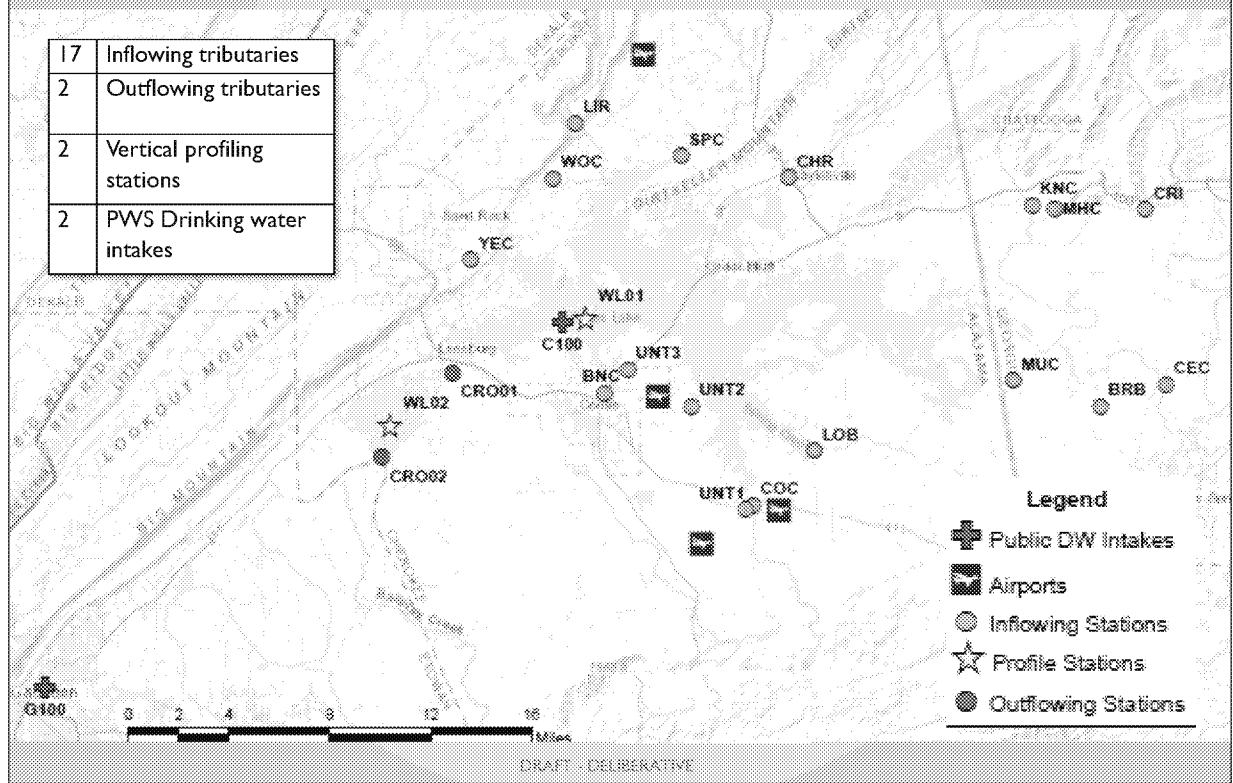
## **Study Objectives:**

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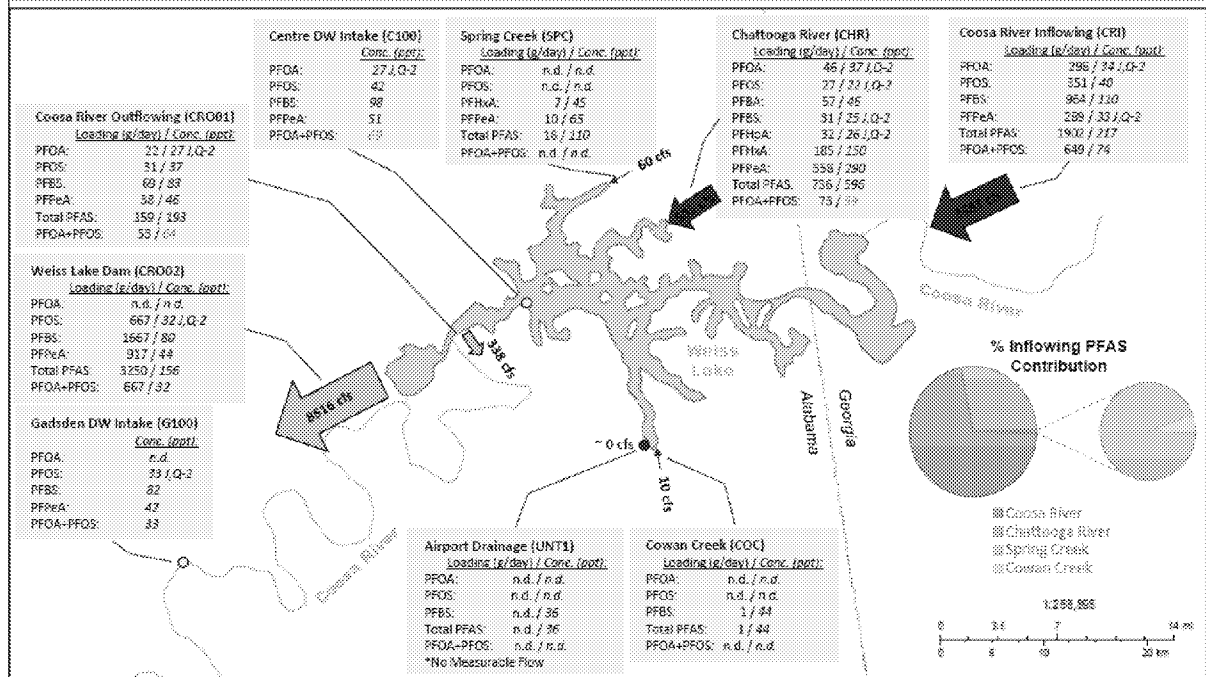
1. Measure background PFAS concentrations and streamflow measurements for all inflowing and outflowing tributary station of Weiss Lake.
2. Estimate mass loading rates of PFASs into Weiss Lake.
3. Examine vertical distribution of PFASs within the lake water column and understand the correlation with water quality parameters.

# Study: Prioritization of PFAS inputs into Lake Weiss

|    |                             |
|----|-----------------------------|
| 17 | Inflowing tributaries       |
| 2  | Outflowing tributaries      |
| 2  | Vertical profiling stations |
| 2  | PWS Drinking water intakes  |

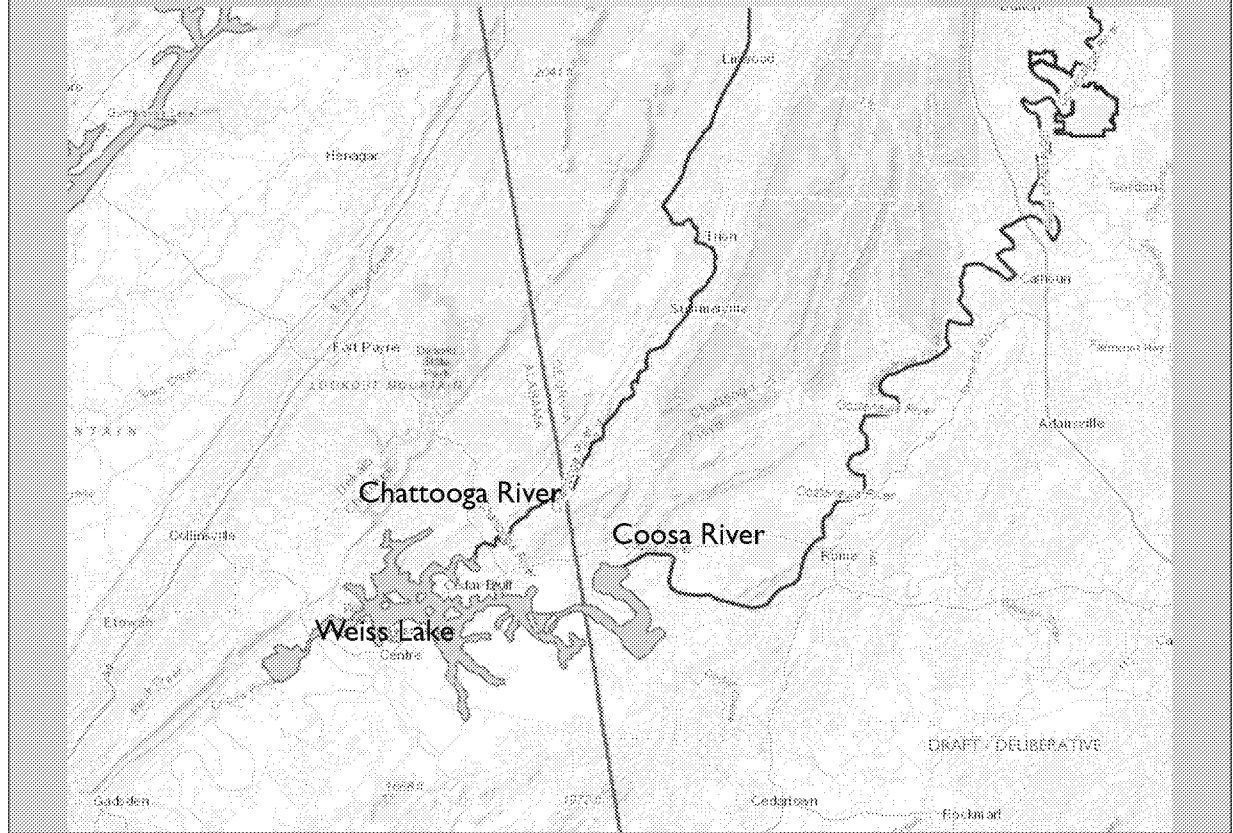


# Results: Prioritization of PFAS inputs into Lake Weiss



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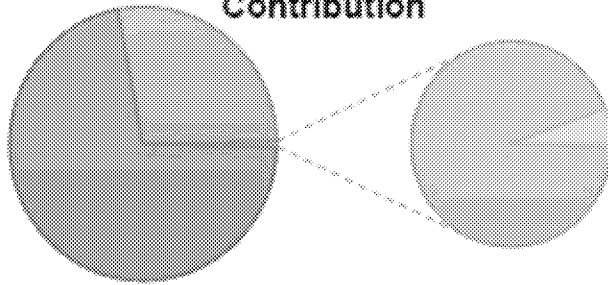
# GA/AL Cross-Jurisdictional Waters



13 co-located SW and sediment samples from LaFayette through Weiss Lake

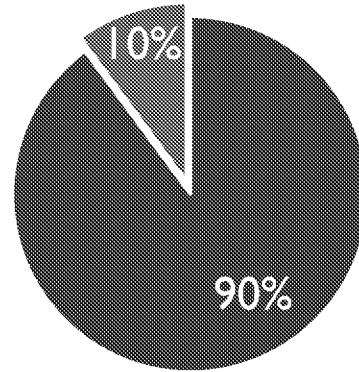
# PFAS CONTRIBUTION

**% Inflowing PFAS Contribution**



■ Coosa River  
■ Chattooga River  
■ Spring Creek  
■ Cowan Creek

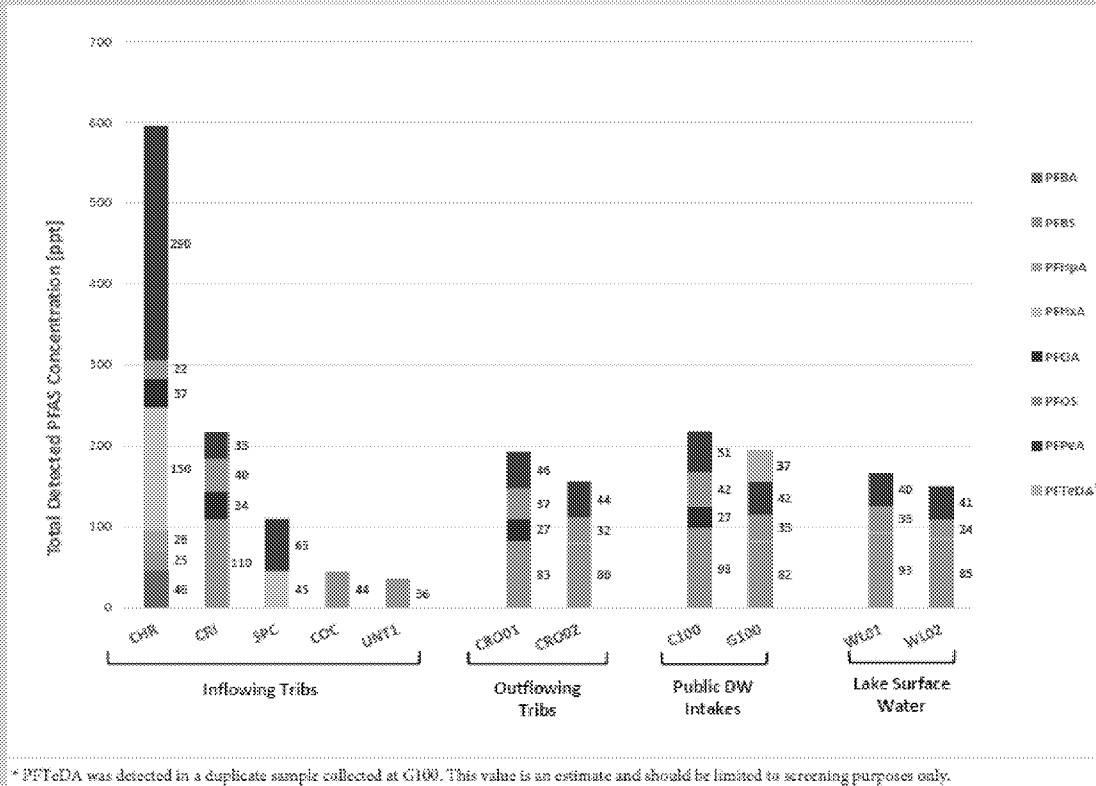
**Mass Loading:  
PFOA/PFOS Contributions to Lake Weiss**



■ Coosa ■ Chattooga

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# Composition of PFAS

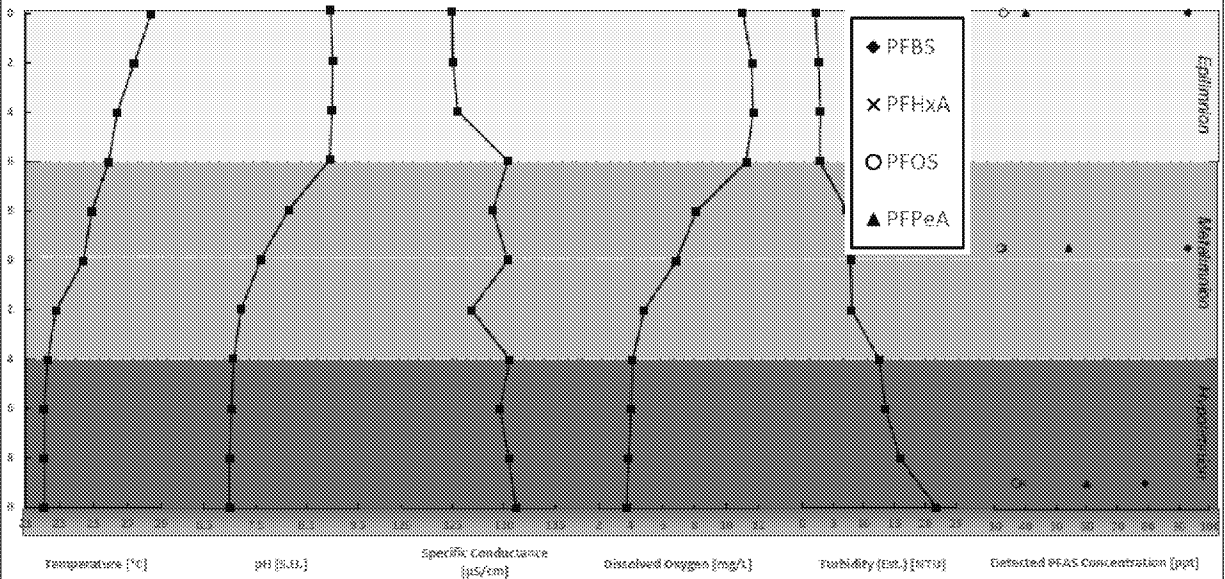


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# VERTICAL PROFILE – WL01

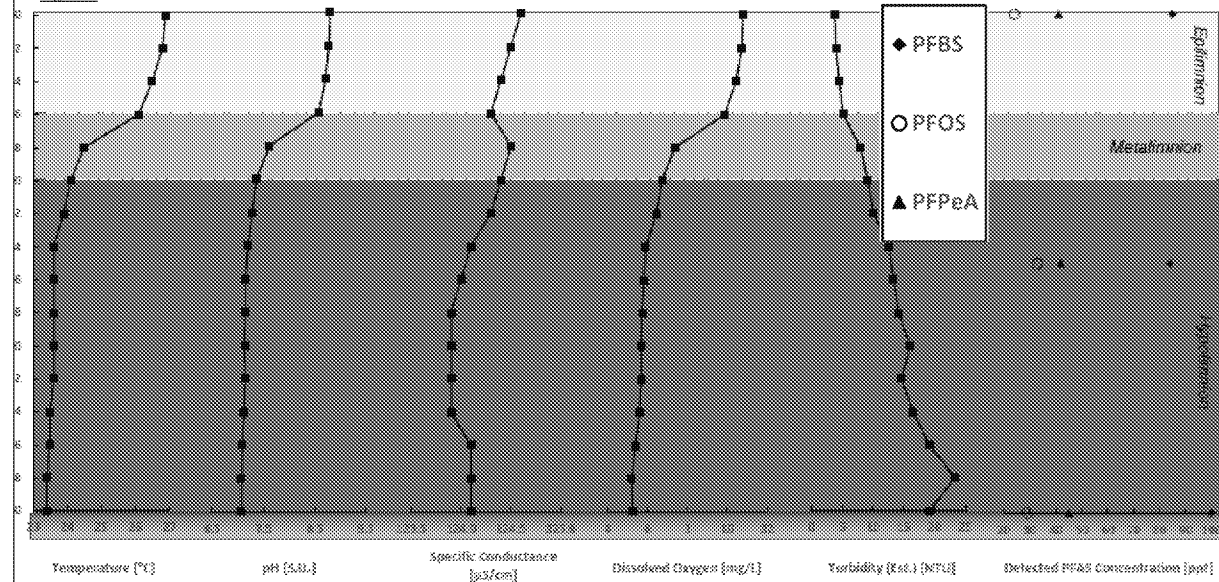
Figure 4: Vertical Profile of WL01



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# VERTICAL PROFILE – WL02

Figure 5. Vertical Profile of WL02



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# **PRIORITIZATION OF PFAS CONTRIBUTIONS TO WEISS LAKE**

## **Conclusions:**

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- Coosa River inlet sample
  - 74 ppt (combined PFOA/PFOS)
  - Largest mass loading rate - 7x flow from Chattooga
  - 90% of combined PFOA/PFOS; 72% of total PFAS
- Chattooga River inlet sample
  - 59 ppt (combined PFOA/PFOS)
  - Greater diversity of PFASs and largest total PFAS concentration
- Consistent distribution of 3 PFASs (PFOS, PFPeA, PFBS) vertically (up to 30 feet below surface) throughout a thermally stratified water column.
  - PFAS contributions from surface water and sediment samples
- Most prevalent PFASs: PFPeA and PFBS (short-chain compounds)

Question: Can we use the LHA as reference to surface water PFOA/PFOS?



# UPPER COOSA RIVER BASIN

**September 2019**

In the past, LAS considered no discharge permit. No NPDES outfall. Even though they monitor for contaminants in the river and have limits. It's a state facility.

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# **ASSESSMENT OF RESUSPENDED SEDIMENTS AS A SOURCE OF PFAS TO THE UPPER COOSA RIVER BASIN**

Conasauga, Oostanaula and Coosa Rivers

## **Study Objectives:**

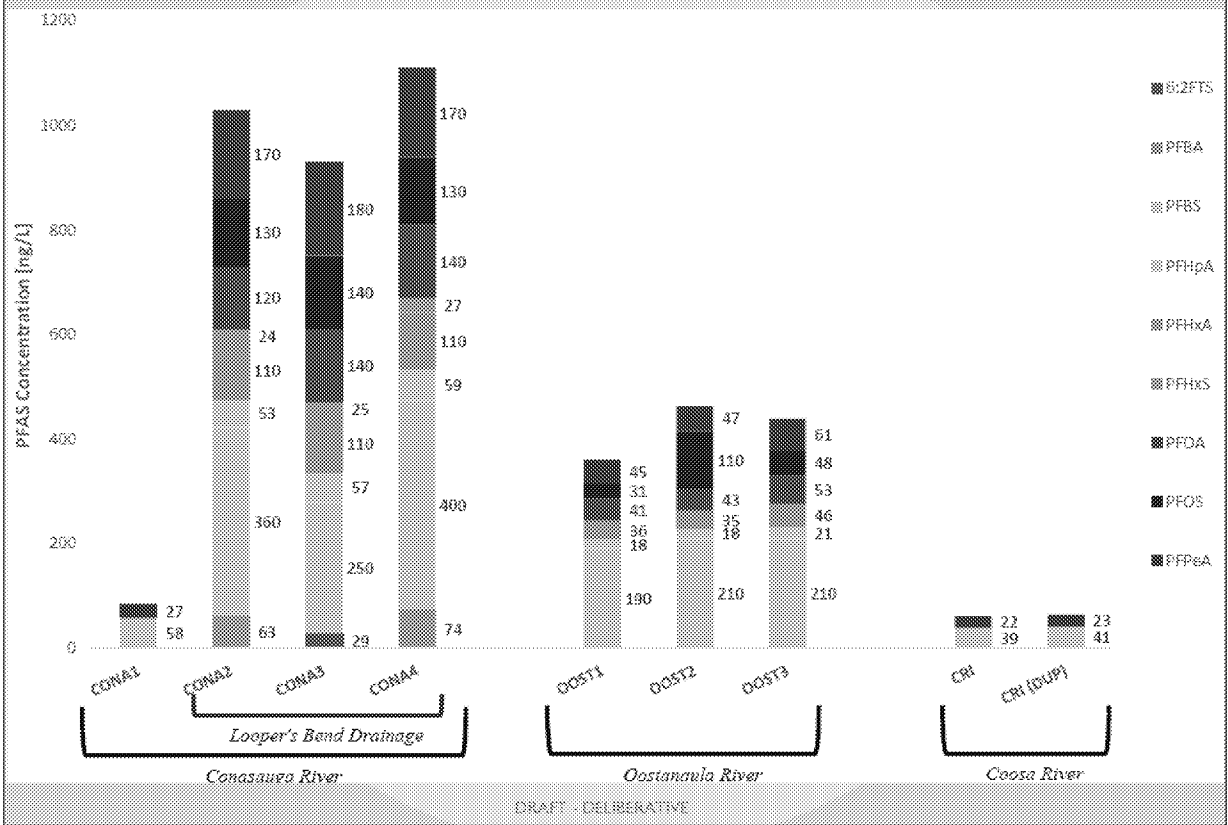
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1. Measure PFAS concentration and composition in stream sediments impacted by the Looper's Bend LAS along the Conasauga into the receiving waters of the Oostanaula and Coosa Rivers.
2. Compare PFAS concentration and composition in co-located surface water and sediment samples.
3. Estimate instantaneous suspended sediment flux to determine the downstream PFAS migration of sediments.

\*Targeted base-flow conditions to avoid PFAS Inputs from surface water runoff and dilution



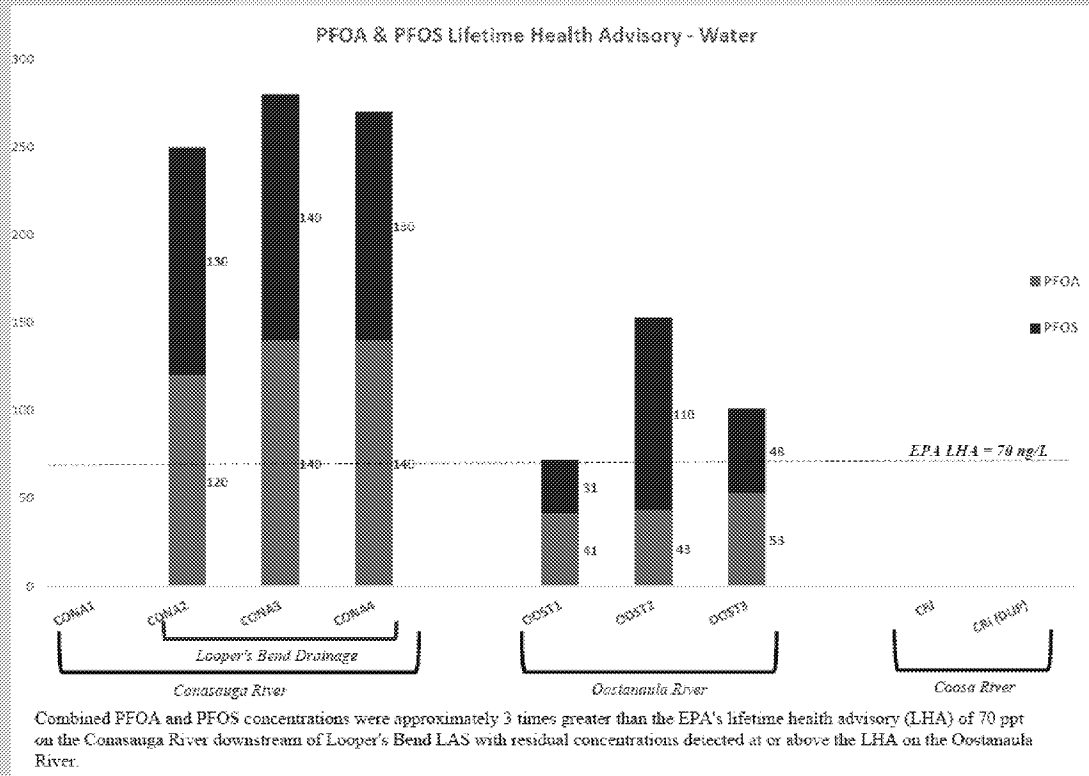
# Composition of Detected PFAS – Surface Water



# Distribution and Mass Loading of PFAS Surface Water



# PFOA and PFOS in Surface Water



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**Legend**

- Sampling Location
- River
- Cooper's Bend LAS

**PFAS Concentration (ng/g dry)**

Legend for bar chart:
 

- PFOS
- ▨ PFOS
- ▩ Other PFAS

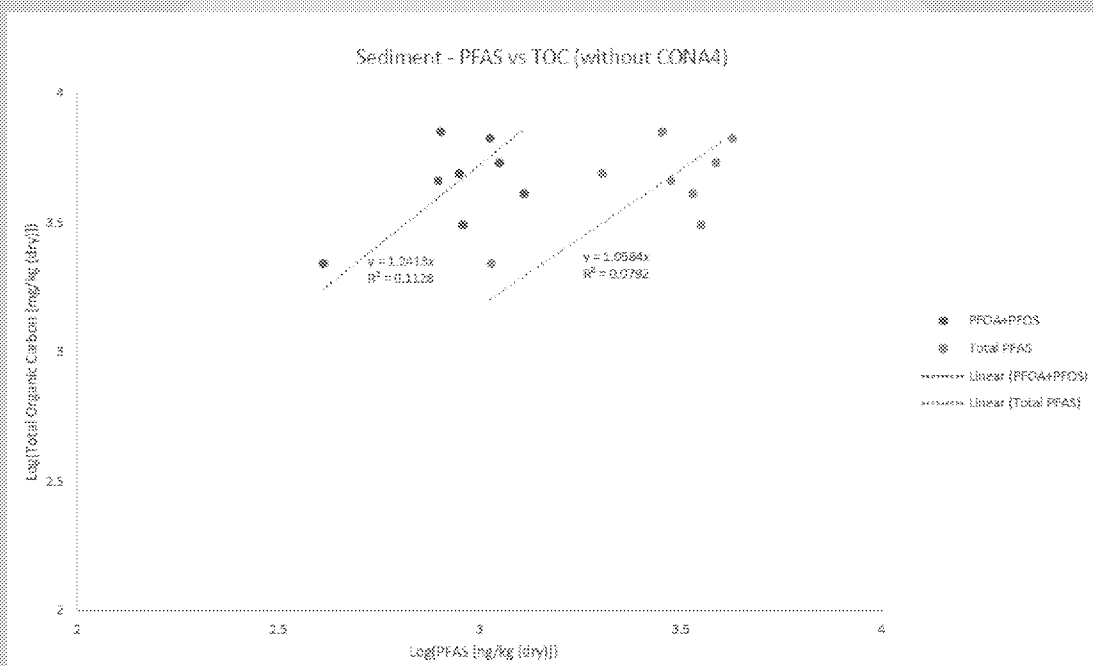
**PFAS Concentration Data (ng/g dry)**

| Compound   | Event 1 (Solid) | Event 2 (Hatched) | Event 3 (Dotted) |
|------------|-----------------|-------------------|------------------|
| PFOS       | ~1500           | ~1500             | ~1500            |
| PFOS       | ~1500           | ~1500             | ~1500            |
| Other PFAS | ~1500           | ~1500             | ~1500            |

All PFAS concentrations are in  $\mu\text{g/kg}$  dry (ppt). Pie charts are shown in relative proportions based on combined PFQA and PFOS.

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# TOC and PFAS Retention



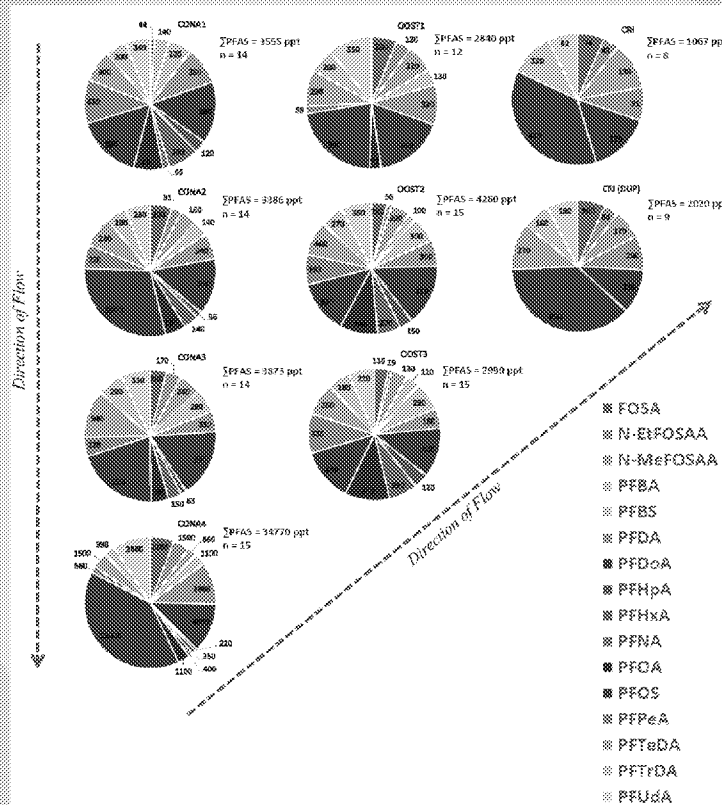
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# Functional Group and Classification

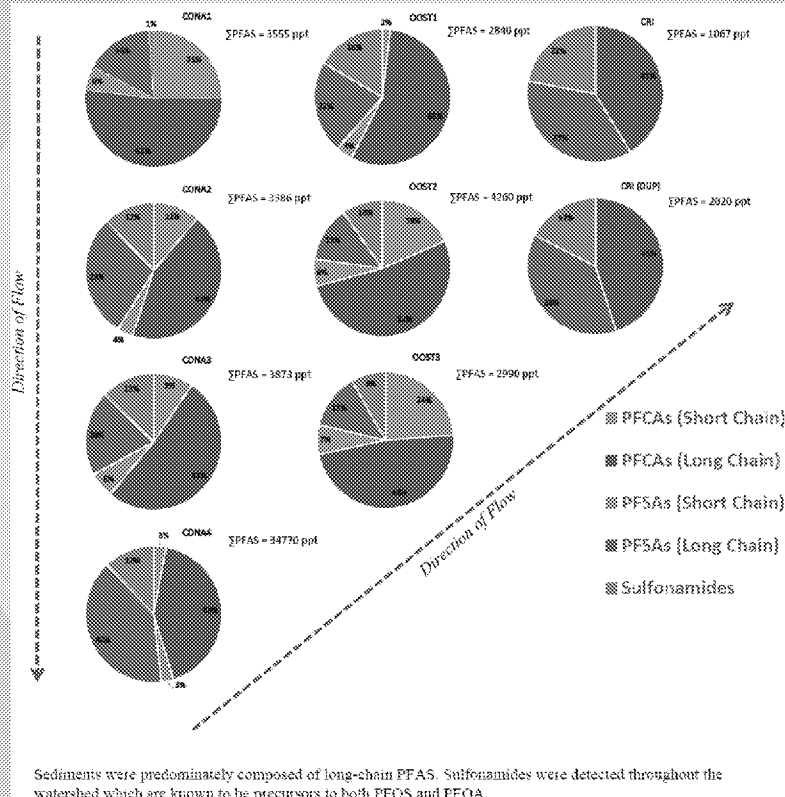
| Carbon Chain Length | PFAS Analyte | Class       | Functional Group |
|---------------------|--------------|-------------|------------------|
| 14                  | PFTeDA       | Long Chain  | PFCAs            |
| 13                  | PFTrDA       |             |                  |
| 12                  | PFDoA        |             |                  |
| 11                  | PFUDA        |             |                  |
| 10                  | PFDA         |             |                  |
| 9                   | PFNA         |             |                  |
| 8                   | PFOA         |             |                  |
| 7                   | PFHpA        | Short Chain |                  |
| 6                   | PFHxA        |             |                  |
| 5                   | PFPeA        |             |                  |
| 4                   | PFBA         |             |                  |
| 10                  | PFDS         | Long Chain  | PFSA's           |
| 9                   | PFNS         |             |                  |
| 8                   | PFOS         |             |                  |
| 7                   | PFHpS        |             |                  |
| 6                   | PFHxS        |             |                  |
| 5                   | PFPeS        | Short Chain |                  |
| 4                   | PFBS         |             |                  |
| 10                  | 8:2 FTS      | Precursor   | Fluorotelomers   |
| 8                   | 6:2 FTS      |             |                  |
| 6                   | 4:2 FTS      |             |                  |
| 10                  | N-EtFOSAA    | Precurscr   | Sulfonamides     |
| 9                   | N-MeFOSAA    |             |                  |
| 8                   | FOSA         |             |                  |
| 6                   | HFPO-DA      | GenX        | Fluoropolyether  |

Classifications adapted from ITRC, 2018

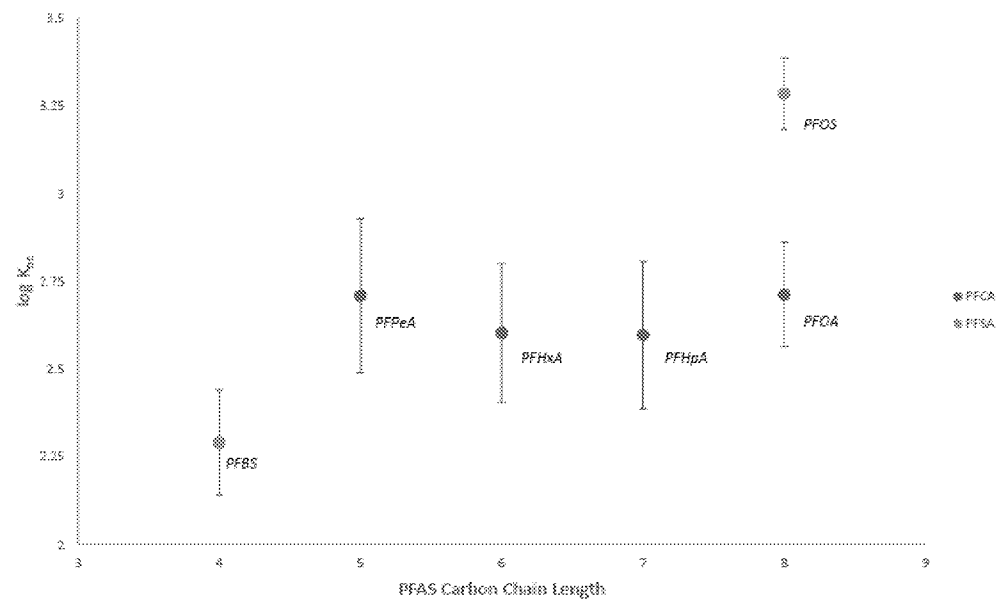
# Composition of Detected PFAS in Sediments



# PFAS Class and Functional Group in Sediments



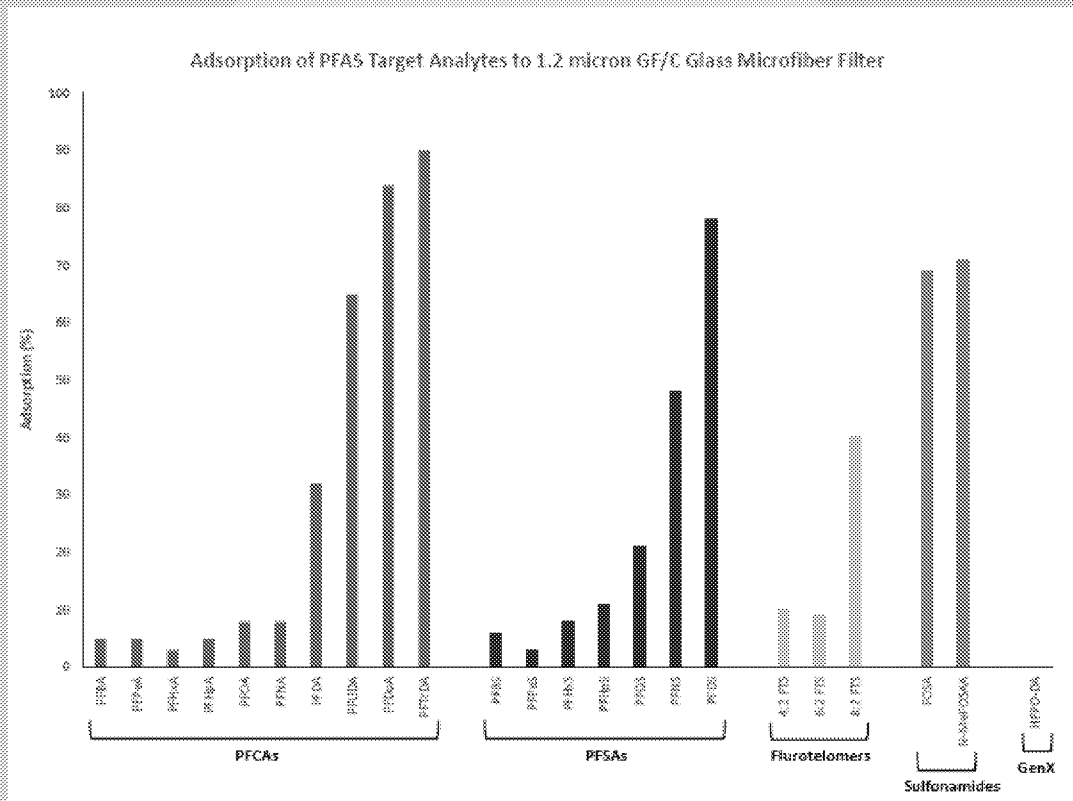
# Organic Carbon on PFAS Retention in Sediments



The effect of organic carbon and the retention of PFAS in sediments is dependent on both the chain length and functional group. PFAS retention in sediments generally increases with increasing chain length and PFSA's (e.g. PFOS) are more likely to be retained compared to PFCA's (e.g. PFOA) of the same chain length.

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# PFAS Adsorption Capacity of Glass Filter



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# UPPER COOSA RIVER BASIN: SURFACE WATER-SEDIMENTS

Conasauga, Oostanaula and Coosa Rivers

## Conclusions:

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- **Surface water:**

- PFBS and PFPeA (short-chain compounds) are most prevalent throughout the watershed (similar to Prioritization Study)
- Generally dominated by short-chain PFSA and PFCAs (highly water-soluble)

- **Sediment:**

- Dominated by long-chain PFCAs, followed by long-chain PFSA and sulfonamido precursors (higher adsorption capacity)
- Total PFAS influenced by the presence of TOC

# UPPER COOSA RIVER BASIN: SURFACE WATER-SEDIMENTS

Conasauga, Oostanaula and Coosa Rivers

## Conclusions:

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- **Mass Loading:**
  - Conasauga River: 74% of PFOA/PFOS Mass Loading to the Oostanaula River
  - Mass loading analysis limited by current Method Detection Limit
- Highest diversity and concentrations of PFAS observed downstream of Looper's Bend persisting in the receiving waters
- The contribution of resuspended sediments was inconclusive due to high variability and adsorption loss of PFAS to filters

A graphic featuring three overlapping circles in shades of gray. A dark gray horizontal band is positioned across the middle of the circles. The text "CHATTOOGA RIVER WATERSHED" is written in white, bold, sans-serif capital letters within this band. Below the band, the text "November 2019" is written in black, bold, sans-serif capital letters.

# CHATTOOGA RIVER WATERSHED

**November 2019**

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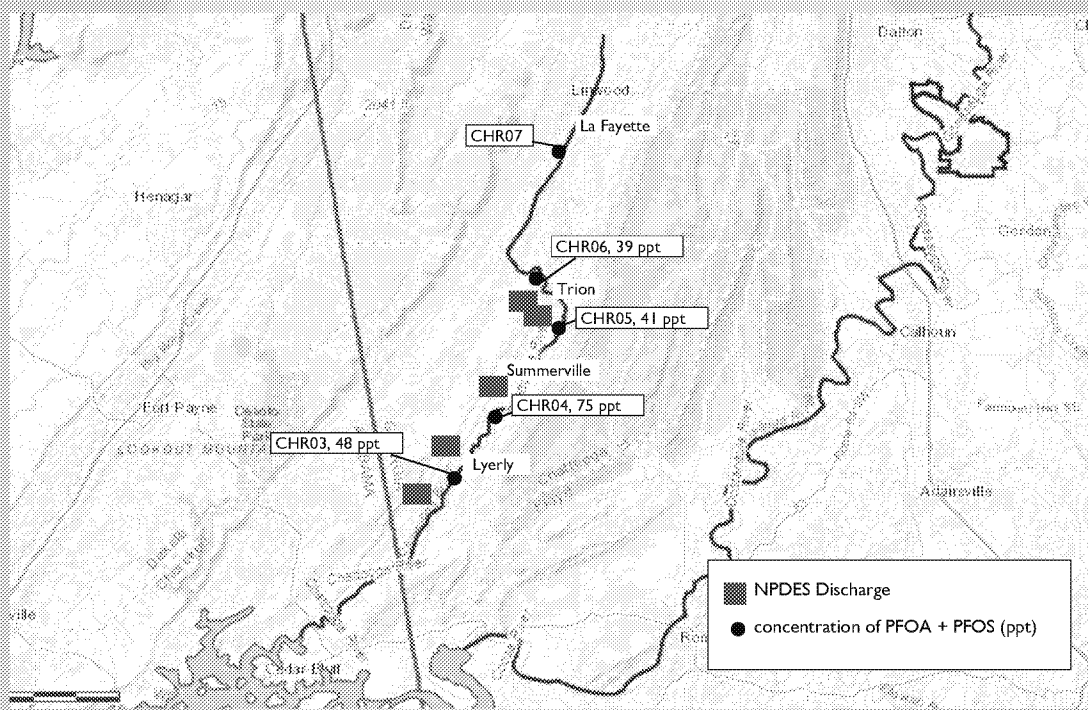
# **Characterization of Ambient PFAS in the Chattooga River Watershed**

## **Study Objectives:**

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1. *Characterize the distribution and instantaneous mass loading of PFAS in the Chattooga River Watershed at near-base flow conditions along key segments determined by the R4 water division.*
2. Collect surface water samples coupled with discharge measurements to compute instantaneous mass loading rates of PFAS along key segments of the Chattooga River Watershed.
3. Collect sediment samples collocated with surface water sample locations to determine the relative distribution and the potential for migration of PFAS contaminated sediments to the receiving waters of

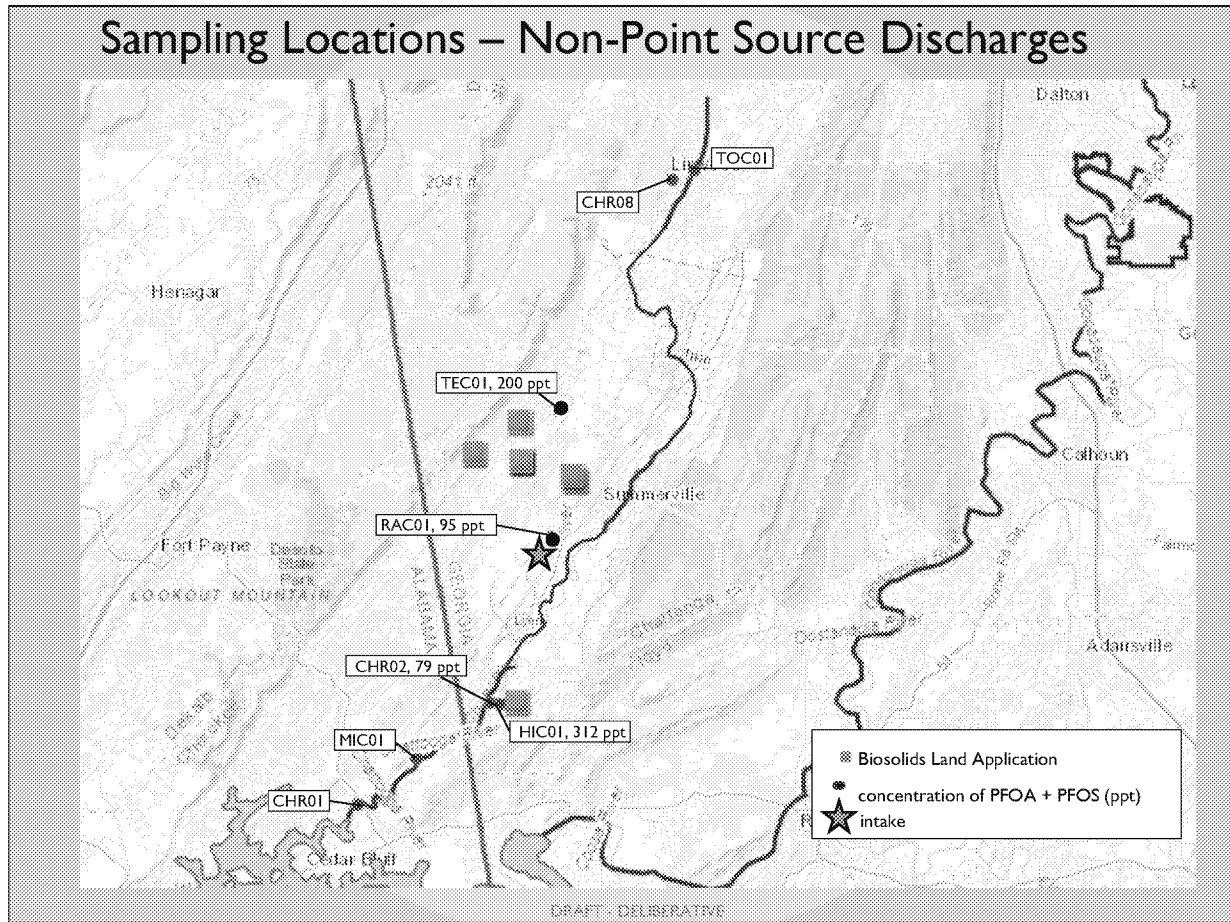
## Sampling Locations – Key Segments



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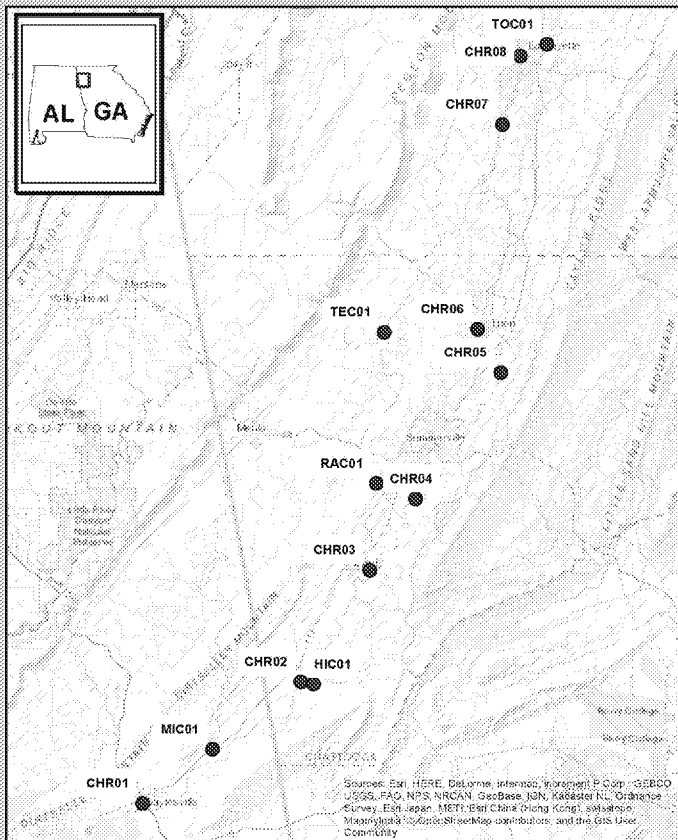
6 SW samples had results above the LHA

## Sampling Locations – Non-Point Source Discharges



Intakes, biosolids fields and SW samples above LHA

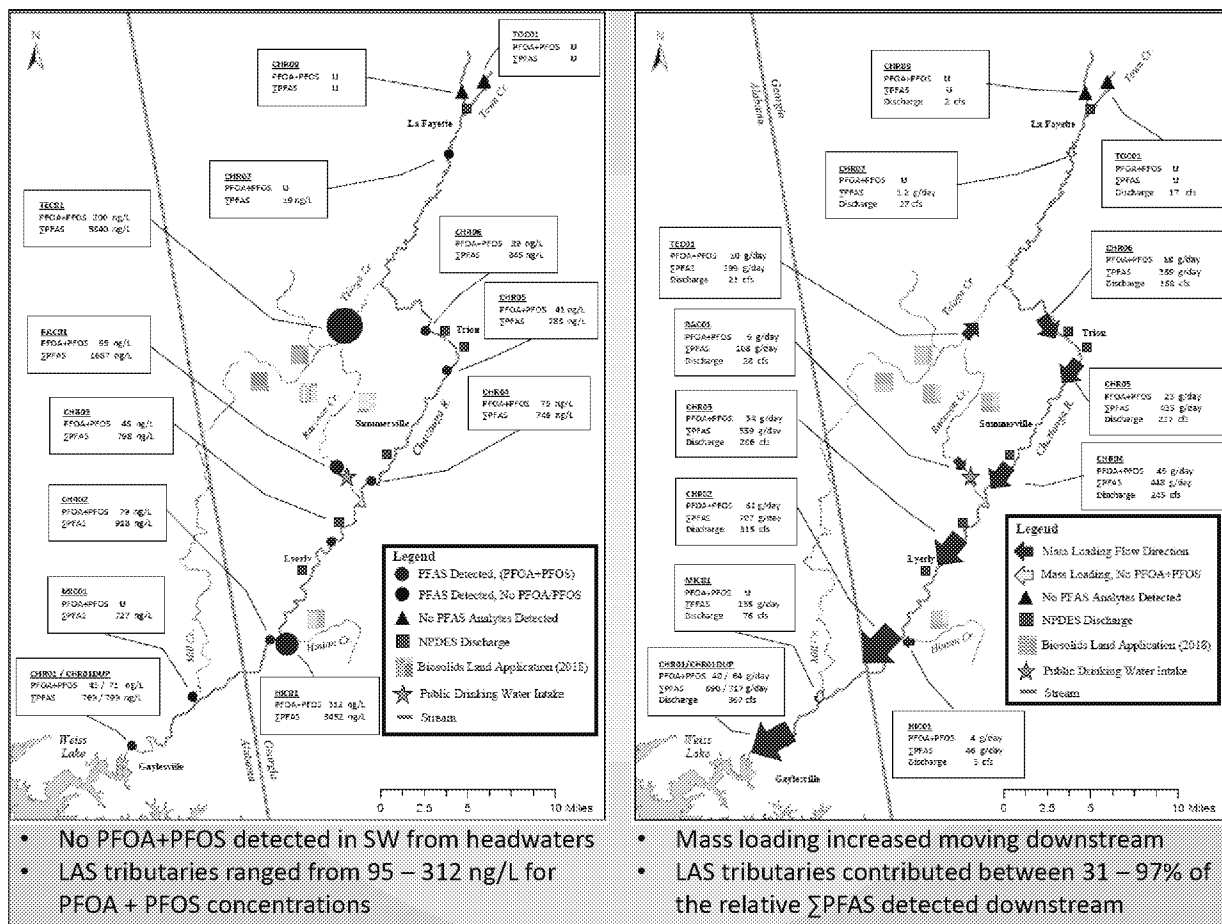
Unlike the Coosa River watershed, more biosolids fields are in the Chattooga



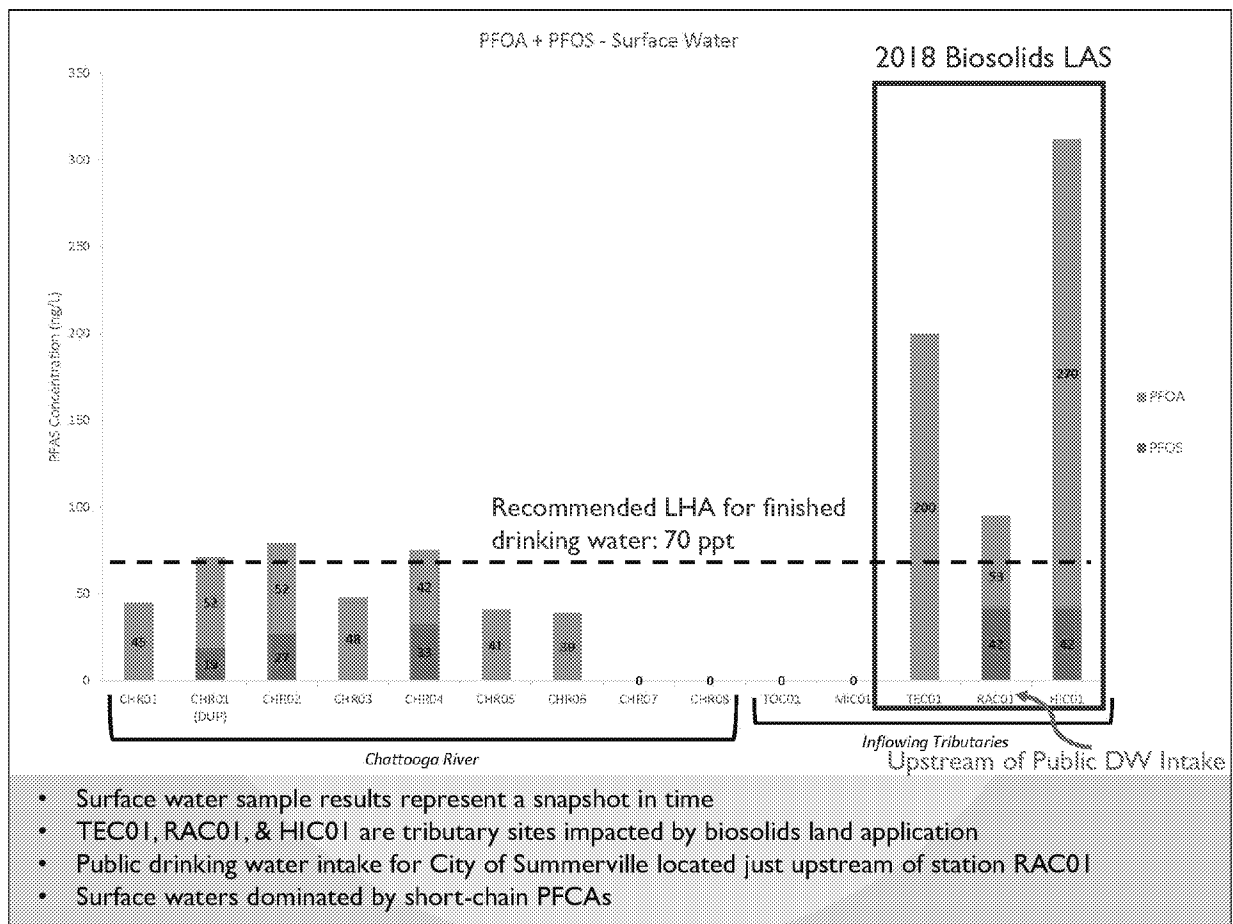
**Total of 13 sites sampled for 25 PFAS Analytes:**

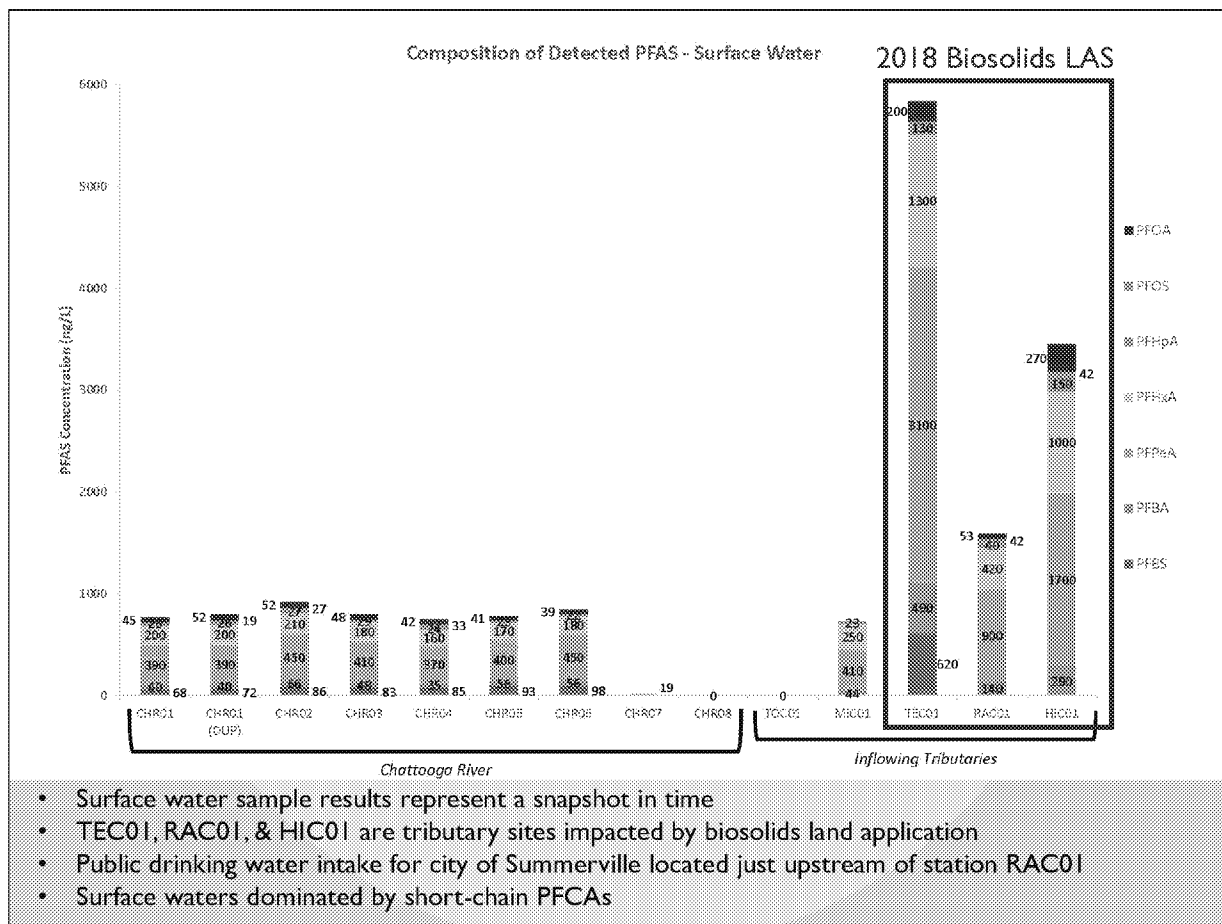
- 8 sites on the main stem
- 3 sites on LAS\* tributaries
- 2 sites from other inflowing tributaries

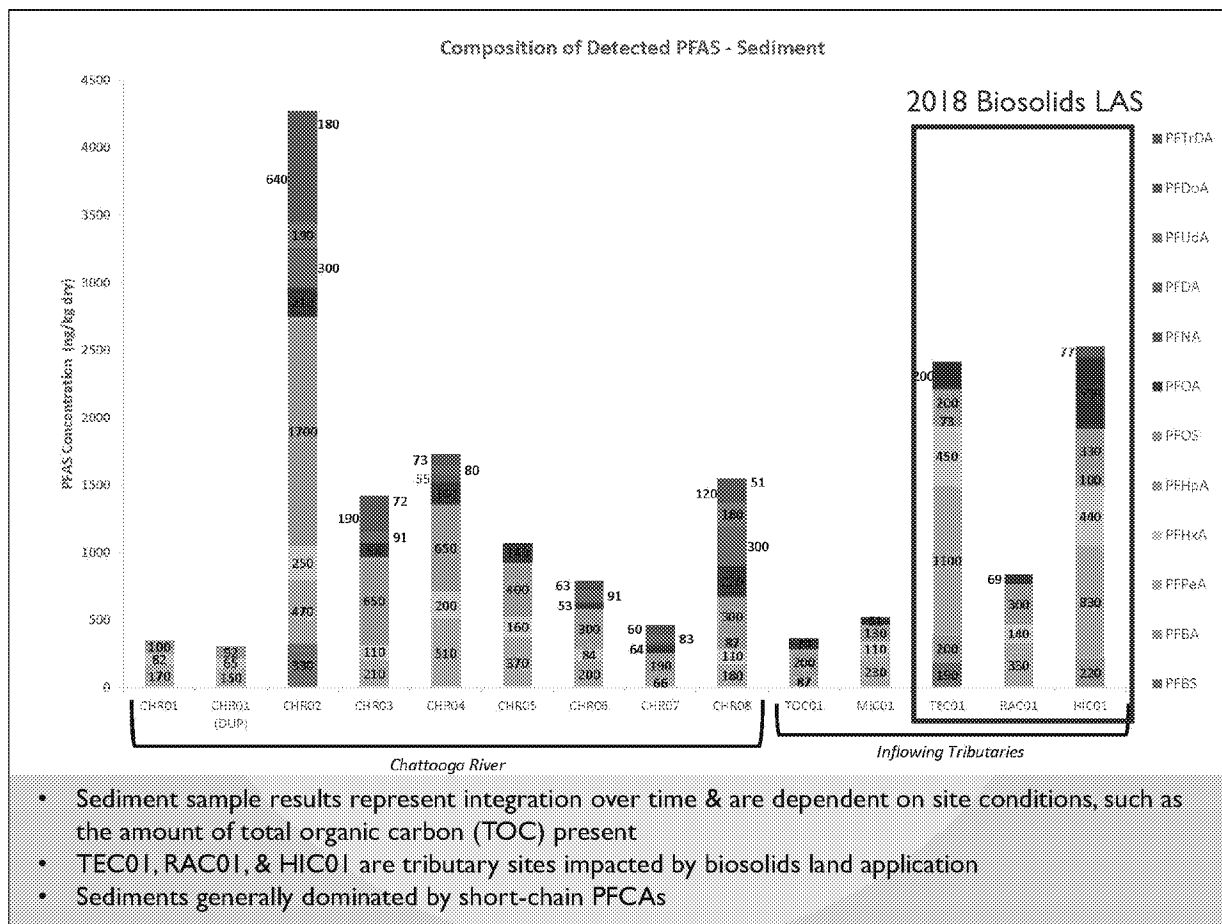
*\*For this presentation, the term LAS refers to biosolids Land Application Sites\**











# Characterization of Ambient PFAS in the Chattooga River Watershed

## Conclusions

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**Sediment Sampling Results:** 12 distinct PFAS compounds detected, majority of samples dominated by short-chain PFCAs

**Surface Water Sampling Results:** 7 distinct PFAS compounds detected, all samples dominated by short-chain PFCAs

- No PFAS detected in surface water samples from headwaters
- Significant contribution of  $\Sigma$ PFAS & combined PFOA+PFOS inputs from tributaries with active biosolids land application sites
- Short-chain PFCAs (C5&C6) dominate both SW & SD, suggesting a local source of short-chain PFCAs &/or their precursors within the watershed
- Transport of both SW & SD contaminated with PFAS from the Chattooga River Watershed → Weiss Lake

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